Leszek Domański, Ewa Zimnoch-Guzowska, Maria Domańska, Henryka Jakuczun, Kazimiera Zgórska, Anna Frydecka-Mazurczyk

Plant Breeding and Acclimatization Institute, Department of Potato Genetics and Parental Lines¹⁾, Research Center Młochów, 05-831 Młochów, Poland, Department of Potato Storage and Processing²⁾, Research Center Jadwisin, 05-140 Serock, Poland

THE DEVELOPMENT OF COLD CHIPPING POTATO PARENTAL LINES FOR BREEDING CULTIVARS SUITABLE FOR PROCESSING

ABSTRACT

The objective of this research was to determine if variation for cold chipping exists in two breeding populations 4×*-*4× and 4×*-*2× potato hybrids. Twenty four potato genotypes of a population of 4×*-*4× hybrids (originated from crosses between parents with good chipping quality) were tested under field conditions at Młochów Research Center of the Plant Breeding and Acclimatization Institute over three years. Traits evaluated included: total tuber yield, marketable yield, starch content, chip colour after cold storage at 4°C, content of reducing sugars, tuber appearance. Out of 24 tested clones three have been selected with outstanding low accumulation of reducing sugars after cold storage (4°C), stable light chip colour and increased starch content. They were also described in trials as having moderate yielding ability, good tuber appearance and satisfied level of resistance to PVY, PLRV and *Phytophthora infestans*. Of four tested diploid parents in 4×*-*2× crosses, the DG 93-332 and the HT/HZ84-PH-151 were more suitable for creation of tetraploid generations combining low content of glucose with increased starch content, good yielding ability and satisfied tuber appearance.

Key words: breeding, cold chipping, potato, reducing sugars, starch content

INTRODUCTION

Most of potato cultivars presently used for the production of potato chips and French fries accumulate significant amounts of reducing sugars when stored at temperatures below 7°C (Burton, 1978; Lisiñska and Leszczyñski, 1989; Zgórska and Frydecka-Mazurczyk, 2000). This cold-induced sweetening process reduces product quality since dark bitter potato chips and French fries result after frying process (Coffin *et al.*, 1987; Sowokinos, 2001). This undesirable flavor and colour of potato chips and French fries is due to products of the Maillard reaction (Schallenberger *et al.*, 1959). Tubers of such cultivars prior to processing must be subjected to reconditioning. Recontitioning at the temperature

Communicated by Henryk J. Czembor

15-18°C for 2-3 weeks of potatoes stored below 7°C decreases the concentration of reducing sugars to acceptable level (Talburt and Smith, 1967; Lisiñska and Leszczyñski, 1989). For potato growers, however, reconditioning requires time and increases their expenses.

At present, an important breeding objective in the development of potato chipping cultivars is resistance to cold-induced sweetening. Such type of cultivars would provide several benefits for raw material quality like: less shrinkage, retention of dry matter, decreased disease loss, limited use of sprout inhibitors and extended marketability (Ewing, 1974; Pereira *et al.*, 1994; Thill and Peloquine, 1995).

Some clones were identified among wild and primitive cultivated potato species which showed little accumulation of reducing sugars after cold storage (Thill and Peloquin, 1994; Jakuczun *et al.*, 1995; Hamernik and Hanneman, 1998). Introgression of this trait from wild or primitive cultivars into *S. tuberosum* required several generations of crossing, backrossing and selection (Thill and Peloquine, 1995). The regular search of clones capable to produce light coloured chips after storage at 4.5°C was begun in 1965 in Minnesota breeding programme (Lauer and Shaw, 1970). The development of processing cultivars and improved parental lines with low level of reducing sugars at cold storage $(4-6sC)$ is being an important objective of breeding programmes both in North America (Pavek, 1987; Thill and Peloquine, 1995) and in Europe (Mackay *et al.*, 1997; Putz, 1999). However, parental value of good cold-chippers ND860-2, ND2221-6 and another ones containing *S. phureja* ancestry are limited because of their short dormancy (Ehlenfeldt *et al.*, 1990). The time taken for the development of cold-chipping cultivars may be shortened if the desired heritable variation can be found within *S. tuberosum* genepool comprising agronomically adapted material.

Our study was undertaken:

- (1) to provide information about variation for chip colour after cold storage (4°C) within a pool of tetraploid selections and tetraploid-diploid hybrids;
- (2) to identify potato tetraploid clones capable to produce light coloured chips after cold storage at 4°C; (3) to evaluate selected tetraploid clones for tuber yield, quality traits, and some important resistance traits.

MATERIAL AND METHODS

Plant material

Two groups of breeding material were used for the study. The first one was represented by 24 clones and four standard cultivars grown in field trials over three years (1999-2001) at Młochów Research Center of the Plant Breeding and Acclimatization Institute (IHAR). The clones had been identified during routine screening in 1998 as having potential to

produce light golden chips after cold storage (4°C) and reconditioning. Selected standard cultivars differed in their suitability for processing. Cv. Saturna is commonly processed into chips both in West Europe and in Poland. Mila and Triada are Polish cultivars both for table and processing use, while Maryna is non-chipping cultivar (Zgórska and Frydecka-Mazurczyk, 1997).

The second group of breeding material consisted of 96 seedlings originated from four interploid crosses (4×*-*2×) between good chipping tetraploid and diploid parents and 48 seedlings from two 4×*-*4× crosses was evaluated in field experiment in 2000 and 2001. Each of 4×*-*2× crosses (Brodick × DG 98-916; Signal × DG 88-89; Signal × DG 93-332; M-62643 \times HT/HZ84-PH-151) and 4×-4× crosses (M-62643 \times Signal; M-62654 \times Signal) was represented by 24 seedlings pre-selected for tuber appearance from the initial 60 individuals.

Each trial was planted as a randomized complete block design (Table 1). Soil at IHAR Młochów's Experimental Field is loamy sand and was fertilized with $100 \text{ kg} \times \text{ha}^{-1}$ of N, 75 kg \times ha⁻¹ of P₂O₅, 165 kg \times ha⁻¹ of K₂O, 75 kg \times ha⁻¹ of S, and $19 \text{ kg} \times \text{ha}^{-1}$ of MgO. Preventive disease and insect control were used in all trials. The trials were harvested at full maturity stage.

The overview of field trials conducted at M³ochów Research Center, IHAR

Group of the breeding material Year of the Number of experiment replicates Number of tested genotypes Size of the plot Evaluated traits: Agronomic Chip colour Sugar content $I \text{ (clones)}$ 1999 2 28 (24 clones + 4 cvs) 8 plants + $I \text{ (clones)}$ 2000 2 28 (24 clones + 4 cvs) 30" $I \text{ (clones)}$ 2001 2 28 (24 clones + 4 cvs) 30' II (seedlings) 2000 1 24×6 families: 4 families $4 \times -2 \times 1$ plant 2 families 4×-4× II (seedlings) 2000 3 24 clones 2 plants assessed using Potato Test Stripes produced by PRECISION LABS

Traits recorded

The overview of recorded traits is shown in the Table 1. Agronomic traits were evaluated in three trials: I/2000, I/2001, and II/2001. Yield of each plot was graded according to tuber size. Weight of all tubers with diameter > 40 mm, expressed as a percent of total yield was rated as marketable yield. Depth of eyes were scored on 1-9 scale, where 9 means very shallow eyes. The incidence of tubers with defects was recorded in percentage according to method applied by Domañski (2001).

Date about resistance of selections to pathogens were obtained in routine testing of parental lines according to methods given by Pietrak

Table 1

(2001) in case of evaluation for resistance to viruses PVY, PLRV and PVM; Stefan and Malinowska (2000) in case of evaluation for resistance to wart disease and eelworm; Zarzycka (2001) in case of evaluations for resistance to late blight and mixed rots caused by *Erwinia carotovora* ssp. *atroseptica* + *Fusarium* sp.

At harvest one sample of 10 medium sized (55-60 mm diameter) tubers, free from disease and damage symptoms was collected from each plot to evaluate chip colour. Chips were produced directly after three months storage at 4°C. Three 1.6 mm thick slices were cut transversally from mid-section of each tuber. Slices were fried in hydrogenated vegetable oil at 180°C in a thermostatically controlled chip fryer until bubbling ceased. Chip colour was visually assessed on a 1 to 9 scale using the colour cards of IBVL $(9 =$ extremely pale).

The 25-tuber samples of nine selections that produced acceptable to industry chips after cold storage were analysed for reducing sugar and sucrose contents using method given by Talburt and Smith (1967).

Each of 144 seedlings from the trial II/2000 and standard cultivar Saturna were assessed for glucose content using Sugar Test Tapes produced by PRESISION LABS, INC. (West Chester, USA). In 2000, after 4 months storage at 4°C, the 4-tuber samples were taken for assessment of glucose content. In 2001 the assessment was repeated in the case of 22 seedlings with significantly lower level of glucose than cv. Saturna. The glucose content evaluation included two separate four-tuber samples from each seedling after their storage for 4 months at 4°C.

Statistical analysis

Combined analysis of variance for chip colour was carried out, next the analysis of individual genotypes, within which genotype \times years interactions were estimated (Table 2). To identify clones with higher chip colour scores than standard cultivar Saturna across years, a pair-wise analysis of variance between individual tested clones and above mentioned standard was additionally carried out. All these analyses were carried out using SERGEN v.3 software (Caliñski *et al.*, 1998). Additionally, twelve ANOVA's, five for glucose content of Ist year seedlings, and one for each of assessed agronomic trait of the $IInd$ year seedlings were performed.

RESULTS AND DISCUSSION

The analysis of variance for chip colour after cold storage $(4^{\circ}C)$ for selected 24 clones from *4×-*4× crosses and four cultivars indicated that the variability of the means for genotypes, years, and genotypes \times years interaction were all highly significant (Table 3). It is worth to point out that the input of genotypic component in the observed phenotypic variation found was high (62.4%). This result has shown that variation for chip colour after cold storage (4°C) exists between tetraploid *S. tuberosum* selections.

Average chip colour of selected tetraploid clones from 4×-4×progenies after three months storage at 4°C and their interaction with years. Three year data, M³ochów 1999-2001

*, ** - significant at α = 0.05 and α = 0.01 respectively; ns – not significant

Table 3

Mean squares and variance components from combined ANOVA for chip colour after three month storage at 4°C directly and after reconditioning for two weeks of 24 tetraploid clones and four standard cultivars. Three year data, M³ochów 1999-2001.

			Mean squares and variance components from combined ANOVA for chip colour after three month storage at 4° C					
	Source of variation	Degrees of freedom		Directly	After reconditioning for two weeks			
			Mean square	Variance components	Mean square	Variance components		
Years		$\overline{2}$	$1.09**$	$0.018 - (1.8)$	$1.06**$	$0.018 - (2.2)$		
Genotypes		27	$4.42**$	$0.627 - (62.4)$	$3.92**$	$0.597 - (74.2)$		
	Genotypes \times Years	54	$0.66**$	$0.300 - (29.9)$	$0.34**$	$0.150 - (18.6)$		
Error		81	0.06	$0.060 - (6.0)$	0.04	$0.040 - (5.0)$		

** - Significant at a = 0.01; proportional values are given in brackets

It was high enough to indicate that genetic improvement for the above mentioned trait could be made through selection. Studies at the Scottish Crop Research Institute also provided evidence for the presence of variation for low-temperature sugar stability among *S. tuberosum* clones. (Brown *et al.*, 1990; Mackay *et al.*, 1990). After storing the 24 clones and four cultivars for three months at 4° C, none of the cultivars (Table 2) chipped acceptably to industry directly from cold storage (with chip colour scored > 6). Four clones M-62705, M-62724, M-62741, M-62774

Table 2

96 *L. Domañski et al.*

produced chips with acceptable colour scored from 6.1 to 6.4. Each of these clones indicated also low interaction with years and their mean scores for chip colour exceeded this one of standard cultivar Saturna, the most important chipping cultivar in Poland and Europe. Above mentioned selections were also stable for starch content (Domañski *et al.*, 2002).

Trait	M-62705	M-62724	M-62741	M-62774	cv. Saturna
Maturity / No of trials	em/2	em/2	m/2	m/2	m/2
Total tuber yield: t/ha	41.5	38.5	46.7	40.5	37.3
Percent of standard	94	87	99	89	82
Marketable yield: (Percent of total yield)	89	80	97	94	77
Starch content - $(\%)$	19.2	17.4	19.1	19.1	17.0
Tuber shape	r - ov	ov	r - ov	ov	r - ov
Tuber size - (scale 1-9)	6.2	5.5	7.2	6.8	5.0
Depth of eyes - (scale 1-9)	7.3	7.0	7.0	7.0	6.4
Incidence of secondary growth $-(\%)$	0.3	2.5	4.2	6.1	13.8
Incidence of growth cracking $-(\%)$	0.6	0.0	0.6	0.0	0.0
Incidence of hollow heart - $(\%)$	5.0	0.0	30.0	0.0	0.0
Incidence of internal rust spot $-(\%)$	3.5	5.0	29.7	0.8	26.7
Content of reducing sugars after storage at 4° C [% fresh weight]	0.06	0.09	0.15	0.07	0.39
Content of sucrose after storage at 4° C [% fresh weight]	0.24	0.17	0.24	0.14	0.36
Resistance for:					
Wart disease	\mathbb{R}	R	\mathbb{R}	S	R
Potato cyst nematodes (Ro1)	S	S	S	S	R
Late blight; foliage / tuber - (scale 1-9)	6/5	3/3	5/6	4/4	4/5
Erwinia spp. $+$ Fusarium sp. $-$ (scale 1-9)	5	$\overline{4}$	3	5	nt
$PVY - (scale 1-9)$	$(8-9)$	$(8-9)$	$(5-6)$	$(8-9)$	$\overline{4}$
$PLRV - (scale 1-9)$	$(5-6)$	$(5-6)$	(4)	(7)	6.5
$PVM - (scale 1-9)$	$(5-6)$	$(7-8)$	(7)	(4)	4

Agronomic characteristics and disease resistance of selections with pale chip colour after cold storage. Agronomic data from 2000 and 2001

Table 4.

Explanation: - for early maincrop (em) trials as standard check were used cvs: Kolia, Mila for maincrop (m) trials as standard check were used cvs: Ania, Danusia; r -ov = round-oval;ov = oval, initial evaluation – given in brackets (), R – resistant, S – susceptible, scale 1-9 (9 = best) nt – not tested

The identification of good chipping clones used as parental lines for further breeding resulted with three selections. The selections M-62705, M-62724, M-62774 originating from parental line breeding program showed little accumulation of reducing sugars and significantly decreased content of sucrose after cold storage in comparison to cv. Saturna, increased starch content and good tuber appearance (Table 4). They

were also described in trials as having moderate yielding ability and satisfied level of resistance to pathogens except that for potato cyst nematodes. The selections M-62705, M-62724, M-62774 besides of good chipping characteristics demonstrated also high resistance to PVY and moderate resistance to PLRV, and PVM. The selections M-62705, M-62724 posses additionally resistance to wart disease and medium resistance to mixed rots. The clone M-62741 is useless as a parent because of the presence of severe internal defects in its tubers.

Table 5

Analysis of variance of four 4×-2× families and two 4×-4× families for content of glucose after 4 months cold storage at 4°C. M³ochów 2000

Source of variation	Degrees of freedom	Mean square	
$4 \times -2 \times$ and $4 \times -4 \times$ families		$0.333**$	
$4\times -2\times$ families		$0.392**$	
$4\times$ -4 \times families		0.074 ns	
$4 \times -2 \times$ families vs. $4 \times -4 \times$ families		$0.420**$	
Error		0.013	

Table 6

Analyses of variance of seedlings within 4x-2x families for content of glucose after 4 months cold storage at 4°C. M³ochów 2000

	Degrees of freedom	Mean squares in families:				
Source of variation		Brodick \times DG 88-916	Signal \times DG 88-89	$Signal \times DG$ 93-332	$M-62643 \times$ HT/HZ84-PH-151	
Seedlings	23	$2.200**$	$1.139**$	$0.937**$	$2.132**$	
Error		0.077	0.118	0.125	0.272	

Analysis of variance (Table 5) indicated highly significant differences between 4×*-*4× and 4×*-*2× families for the level of glucose after 4 months of cold storage. The majority of the variation observed was due to differences among 4×*-*2× families. Appreciable variation (Table 6) was present among seedlings within 4×*-*2× families as well. Both types of variation existed among 4×*-*2× progenies, providing the opportunity for selection of improved clones for use in further cycles of crossing.

Average agronomic performance of seedlings selected for low content of glucose, indirect measure of acceptable chip colour, was presented in Table 7. The higher frequency of perspective clones was observed in progenies derived from tetraploid-diploid crosses than tetraploid crosses. During selection following criteria were applied: low accumulation of glucose (< 0.25% of fresh weight); yield, mean tuber weight, and depth of eyes (\geq mean of cv. Mila – LSD_{0.05}); freedom from tuber defects (secondary growth, growth cracking, short dormancy). Out of tested diploid parents, the DG 93-332 and the HT/HZ84-PH-151 were more suitable for creation of progenies combining low content of glucose

98 *L. Domañski et al.*

with increased starch content, yielding ability and satisfactory tuber appearance. However, clones derived from HT/HZ84-PH-151 (containing *S. phureja* ancestry) often produced tubers with short dormancy.

Table 7

Seedling	Half-sib family	Tuber vield [kg per 2 plants]	Mean tuber weight [g]	Starch content $[\%]$	Approximate glucose content $[1-5 \text{ score}]$	Depth of eyes $[1-9 \text{ score}]$	Com- ments
$R-5$	DG 98-916 $(2\times)$	2.02	81.0	13.4	2.3 ab	5.8	
$R-15$	DG 98-916 $(2\times)$	1.98	93.4	12.8	2.2 _b	6.8	
$R-16$	DG 98-916 $(2\times)$	2.01	121.6	10.9	1.6 _b	5.9	
$R-45$	DG 88-89 $(2\times)$	2.85	83.8	14.6	2.4 ab	6.9	
$R-56$	DG 93-332 $(2\times)$	2.33	83.9	17.1	1.8 _b	6.4	
$R-57$	DG 93-332 $(2\times)$	2.78	92.6	16.9	2.0 _b	6.8	
$R-58$	DG 93-332 $(2\times)$	2.44	95.8	19.4	2.0 _b	6.7	
$R-60$	DG 93-332 $(2\times)$	3.37	77.1	18.3	2.6ab	6.1	
$R-70$	DG 93-332 $(2\times)$	2.56	77.4	17.6	2.6ab	6.4	
$R-72$	DG 93-332 $(2\times)$	2.81	76.5	16.1	2.4 ab	6.4	
$R-73$	$HT/HZ84-PH-151 (2\times)$	2.13	53.9	16.9	2.2 _b	6.8	sh. dorm.
$R-74$	$HT/HZ84-PH-151 (2\times)$	2.95	62.8	16.4	1.5 _b	7.0	sh. dorm.
R-78	$HT/HZ84-PH-151 (2\times)$	2.67	94.4	17.8	2.6ab	7.5	
$R-79$	$HT/HZ84-PH-151 (2\times)$	3.24	98.0	14.6	2.2 _b	6.7	sh. dorm.
R-82	$HT/HZ84-PH-151 (2\times)$	2.04	78.2	15.8	1.9 _b	6.8	
$R-85$	$HT/HZ84-PH-151 (2\times)$	2.79	61.8	15.6	2.5 ab	6.8	sh. dorm.
R-88	$HT/HZ84-PH-151 (2\times)$	3.37	68.4	16.2	2.3 ab	7.1	
R-91	$HT/HZ84-PH-151 (2\times)$	3.06	63.8	15.3	1.6 _b	7.2	
R-94	$HT/HZ84-PH-151 (2\times)$	2.50	52.1	18.1	1.7 _b	7.1	
$R-110$	$M-62643(4\times)$	2.95	73.1	14.8	1.8 _b	6.4	
Mila		2.03	91.2	17.3		6.6	
Saturna					3.4a		
	LSD.	0.51	23.7	1.2	0.6	0.5	

Agronomic characteristics of seedlings selected for low content of glucose and originated from interploid (4×-2×) crosses. M³ochów 2001

¹ – 0%, 2 - 0.10%, 3 – 0.25%, 4 – 0.50%, 5 – 2.00% glucose in fresh weight, mean of 2000 and 2001; ²Comparable means followed by the same letter are not significantly different (α = 0.05) according to Tukey's Honestly Significant Difference Test; sh. dorm. – short dormancy

The present pool of parents for the development of cold chipping clones, obtained from interploid crosses can be enriched by our selections. However, they need evaluation of agronomic performance and chipping quality in frying tests.

In 2002 our best cold chipping selections the M-62724 and the M-62774 were delivered as parental forms to Polish breeding companies PMHZ Strzekęcin and HZ Zamarte. At this moment the information on their combining ability is not available.

Broad-sense heritability of chip colour after cold storage estimated by Pereira *et al.* (1995) in two families of cold chipping clone ND860-2 was moderate and ranged from 0.39 to 0.66. The clone ND860-2 was found in the origin of cultivar NorValley outstanding in cold-sweetening resistance bred by Novy *et al.* in 1998.

In our opinion parental lines the M-62724 and the M-72774 give a chance to develop new cultivars suitable for processing from cold storage. More, as the parental lines M-62724 (cross: Agria \times DG.88-89) and M-62774 (cross: Agria \times PW-363) posses in their origin parents with positive general combining effects (Domañski *et al.*, 2000).

CONCLUSIONS

A considerable genetic variation in potato chip colour after cold storage exists within the pool of tetraploid parental lines developed at Research Center Młochów of the Plant Breeding and Acclimatization Institute. Best selections have been offered to Polish breeders.

Three selections were identified directly from cold storage (M-62705, M-62724, M-62774) combining low concentration of reducing sugars after cold storage, increased starch content, satisfactory resistances to viruses and good tuber appearance. They were also capable of produce acceptable by industry light coloured chips with reletive high stability of this trait over years.

The use of *2n* polen-producing diploid parents the DG 93-332 and the HT/HZ84-PH-151 may be useful for the development of new cold chipping parental lines. The perspective clones combining low content of glucose with increased starch content, good yielding ability and satisfied tuber appearance were selected from 4×*-*2× progenies with higher frequency than from 4×*-*4× progenies.

ACKNOWLEDGEMENTS

This research was supported in part by Polish breeding companies: HZ Zamarte and PMHZ Strzekecin.

REFERENCES

Brown J., Mackay G.R., Bain H., Griffith D.W., Allison M.J. 1990. The processing potential of tubers of the cultivated potato, *Solanum tuberosum* L., after storage at low temperatures. 2. Sugar concentration. Potato Research 33: 219-227.

Burton W.G. 1978. The physics and physiology of storage. In: Harris P.M. (ed), The Potato Crop. Chapman and Hall, London: 545-601.

Caliñski T., Czajka S., Kaczmarek Z., Krajewski P., Siatkowski I. 1998. SERGEN 3 User's Guide. Institute od Plant Genetics, Polish Academy of Sciences, Poznañ; Department of Mathematical and Statistical Methods, Agricultural University, Poznañ, Poland: 1-53.

Coffin R.H., Yada R.Y., Parkin K.L., Grodzinski B., Stanley D.W. 1987. Effect of low temperature storage on sugar concentrations and chip color of certain processing potato cultivars and selections. J. Food Sci. 52: 639-645.

Domañski L., Zimnoch-Guzowska E., Sieczka M.T. 2000. Estimation of general combining ability at the early stage of potato breeding. Plant Breeding and Seed Science 44: 87-94.

- Domañski L. 2001. Assessment of external and internal defects of potato tubers. In: Zimnoch-Guzowska E., Syller J., Sieczka M. (eds), The methods of evaluation and selection applied in potato research and breeding. Monografie i Rozprawy Naukowe 10a/2001. Plant Breeding and Acclimatization Institute, Radzików, Poland: 101-103.
- Domañski L., Zimnoch-Guzowska E., Paczkowska M. 2002. Preliminary evaluation of the stability of some processing quality traits in frame of the development of parental lines for breeding of potato cultivars for chip industry. Biul. IHAR (in print).
- Ehlenfeldt M.K., Lopez-Portilla D.F., Boe A.A., Johansen R.H. 1990. Reducing sugar accumulation in progeny families of cold chipping potato clones. Am. Potato J. 67: 83-91.

Ewing E.E. 1974. Potato chip colour. Amer. Potato J. 51: 242-243.

Hamernik A.J., Hanneman JR. R.E. 1998. Breeding 2× haploid × species potatoes that chip from 2C cold storage. Amer. J. of Potato Research 75, 6: 278-279.

Jakuczun H., Zgórska K., Zimnoch-Guzowska E. 1995. An investigation of the level of reducing sugars in diploid potatoes before and after cold storage. Potato Research 38: 331-338.

- Lisiñska G., Leszczyñski W. 1989. Potato Science and Technolory. Elsevier Applied Science, London and New York.
- Mackay G.R., Brown J., Torrance J.W. 1990. The processing potential of tubers of the cultivated potato, *Solanum tuberosum* L., after storage at low temperature. 1. Fry colour. Potato Research 33: 211-218.
- Mackay G.R., Tod D., Bradshaw J.E., Dale M.F.B. 1997. The targeted and accelerated breeding of potatoes. Scottish Crop Research Institute. Annual Report 1996/97:40-45.
- Novy R.G., Secor G.A., Farnsworth B.L., Lorenzen J.H., Holm E.T., Preston D.A., Gudmestad N.C., Sowokinos J.R. 1998. NorValley: a white-skinned chipping cultivar with cold-sweetening resistance. Amer. J. of Potato Res. 75: 101-105.

Pavek J.J. 1987. Some interesting aspects of recent and expected developments in potato breeding in North America. Acta Horticulturae 213: 61-65.

- Pereira A.S., Tai G.C.C., Yada R.Y., Coffin R.H., Souza-Machado V. 1994. Potential for improvement by selection for reducing sugar content after cold storage for three potato populations. Theor. Appl. Genet. 88: 678-684.
- Pereira A.S., Tai G.C.C., Yada R.Y., Coffin R.H., Souza-Machado V. 1995. Genetic advance for chip colour in potatoes. Euphytica 84: 133-138.
- Pietrak J. 2001. Estimation of resistance to viruses in Polish breeders selections and foreign cultivars in statutory trials. In: Zimnoch-Guzowska E., Syller J., Sieczka M. (eds), The methods of evaluation and selection applied in potato research and breeding. Monografie i Rozprawy Naukowe 10a/2001. Plant Breeding and Acclimatization Institute, Radzików, Poland: 30-32.
- Putz B. 1999. First potato varieties for the chip industry suitable to be stored at 4°C.In: Abstracts of Conference Papers, Posters and Demonstrations 14th Trennial Conference of the EAPR, Sorrento, Italy, May 2-7 1999. Assessorato, Agricoltura Regione Campania: 333-335.

Shallenberger R.S., Smith O., Treadway R.H. 1959. Role of sugars in the browning reaction of potato chips. J. Agric. Food Chem. 7: 274-277.

Sowokinos J.R. 2001. Biochemical and molecular control of cold-induced sweetening in potatoes. Amer. J. of Potato Research 78: 221-236.

Stefan K., Malinowska E. 2000. Metodyka badania odporności materiałów hodowlanych ziemniaka na raka, mątwika ziemniaczanego i mątwika agresywnego. Ziemniak Polski 2000: 26-30.

Talburt W.F., Smith O. 1967. Potato processing. AVI Publishing Company Inc., Connecticut, USA. Thill C.A., Peloquin S.J. 1994. Inheritance of potato chip color at the 24-chromosome level. Am.

- Potato J. 71: 629-646.
- Thill C.A., Peloquin S.J. 1995. A breeding method for accelerated development of cold chipping clones in potato. Euphytica 84: 73-80.
- Zarzycka H. 2001. Evaluation of potato resistance to some fungal and bacterial diseases. In: Zimnoch-Guzowska E., Syller J., Sieczka M. (eds), The methods of evaluation and selection applied in potato research and breeding. Monografie i Rozprawy Naukowe 10a/2001. Plant Breeding and Acclimatization Institute, Radzików, Poland: 75-89.
- Zgórska K., Frydecka-Mazurczyk A. 1997. Przydatność odmian do przetwórstwa spożywczego. Ziemniak Polski 4: 20-23.
- Zgórska K., Frydecka-Mazurczyk A. 2000. The influence of weather conditions during growing season and storage temperature on processing quality of potatoes. Biul. IHAR 213, 239-251.

Lauer F., Shaw R. 1970. A possible genetic source for chipping potatoes from 40°F storage. Amer. Potato J. 47: 275-278.